INCIDENCE OF HISTAMINE IN MARINE FISHES SOLD IN RETAIL MARKETS IN RELATION TO THEIR CONTENT OF HISTIDINE DECARBOXYLATING BACTERIA*

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ABSTRACT

Histamine contents in 10 species of marine fish collected from retail markets in Cochin were estimated along with their bacterial profile in general and content of histidine decarboxylating bacteria in particular. The study reveals that none of the fish samples surveyed contained histamine above 1.3 mg/100 gm fish which is much below the recommended level as per U.S.F.D.A. specification, even though the histidine decarboxylating bacterial counts were of the order of 10⁴ per gm of fish muscle.

INTRODUCTION

HISTAMINE is a toxic degradation product of the amino acid histidine. Scombroid fishes such as tuna and mackerel and certain nonscombroid fishes possess large amounts of free histidine in their muscle tissues. Histamine formation in fish and fish products is brought about by the bacterial enzyme, histidine decarboxylase and to a lesser extent by autolytic degradation (Kimata, 1961; Anon., 1987).

Considering the severity of histamine poisoning, regulatory measures in fish and fish products have been imposed by countries such as Canada, Federal Republic of Germany, U.S.A., Finland, Denmark and Sweden. Table 1 shows the regulatory limits for histamine prescribed by these countries.

Even though extensive work on histamine formation in scombroid and non-scombroid fishes have been carried out in many countries, only limited work has been reported from India. In the present studies an attempt has been made to investigate how far the fishes

sold through the retail outlets in India are safe with respect to histamine content as well as their content of histidine decarboxylating bacteria.

TABLE 1. Regulatory limits for histamine in fish and fishery products

Country	L	Limit of histamine mg/100 g fish					
U.S.A. (for albacore, skipjack, yellowfin tuna							
and mahi-mahi)	••	20 max. level					
		50 Hazard action level					
Canada	••	10 Representative of decomposition					
F.R. Germany.	••	20 Representative of decomposition					
Denmark	••	30 Hazard action level					
Sweden		20 Hazard action level					

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MATERIALS AND METHODS

Marine fishes were collected from the retail markets in Cochin. The fish muscle was analysed in the laboratory for histamine content and their bacterial profile including histidine decarboxylating bacteria. Histamine was estimated by the method of Hardy and Smith (1976) using spectromic 21 spectrophotometer for measuring the absorbance.

The total viable bacterial count (TPC) was determined by using tryptone glucose agar (TGA). The plates were incubated at $28 \pm 2^{\circ}$ C (RT) for 48 hours and counts taken. The total coliforms were determined using Desocycholate lactose agar (DLA), *Escherichia col*₁ using Tergitol-7 agar, faecal streptococci using KF agar and coagulase positive staphylococci using Baird-Parker agar (FDA, 1973; Difco, 1971). Histidine decarboxylating bacteria (HDB) was determined by direct plating of the fish tissue with tryptone-yeast extract-histidine agar (TYMA) of Neven *et al.* (1981).

RESULTS AND DISCUSSION

All the fish species analysed were more or less in prime condition and none had a bacterial count of more than a million/gram muscle. Detailed bacterial quality of these fishes are presented in Table 2.

The histamine contents of the individual fishes, in relation to their histidine decarboxylating bacteria counts are presented in Table 3. The population of histidine decarboxylating bacteria in the muscle of these fishes varied from 40/g in seerfish to 1.12×10^4 /g in Malabär herring. But the content of histamine in the flesh of these fishes was less than 1 mg/100 g in almost all samples analysed. Further, no direct correlation was observed between the histamine content in fish muscle and their population of histidine decarboxylating bacteria.

The histamine content of all the fish samples surveyed were very small, less than 1.3 mg/ 100 g fish muscle which is quite insignificant to it cause any histamine poisoning to the consumer.

Fish	TPC per g	Total coliforms per g	E. coli per g	Coagulase +ve staphylo- cocci per g	Faecal streptococci per g
Jewfish (Otolithus spp.)	7.64 × 10 ⁴	7.85 × 10 ^s	Nil	208	9.03 × 10 ^a
Indian mackerel (Rastrelliger kanagurta)	6,93 × 10 ³	$7.88 \times 10^{\circ}$	27 7	Nil	4.67 × 10 ^a
Carangid (Chorinemus spp.)	4.30 × 10 ⁴	$6.00 imes 10^{s}$	55	3 × 10*	2.50 × 10ª
Malabar herring Thrissocles spp.	1.39 × 10°	1.03×10^{4}	1.8 × 10*	47	3.82 × 10*
Silverbelly (Lelognathus spp.)	4.75 × 10*	2.90 × 10 ^s	583	4.25 × 10 ^s	8.17 × 10 ³
Kilimeen (Nemipterus japonicus)	1.95 × 10 ⁶	2.48 × 104	522	Nil	5.56 × 10ª
Seerfish (Scomberomorus spp.)	5,50 × 104	1.90×10^{4}	Nil	17	47
Carangid	1.17 × 10 ⁵	1.14 × 10ª	200	9 0	2×10^{a}
Lactarius (Lactarius lactarius)	1.102×10^4	7.76 × 10*	Nil	Nil	1.29 × 10*
Ribbonfish (Trichiurus spp.)	4.7 × 10 ⁶	5,30 × 10*	18	$1.4 imes 10^{s}$	400

TABLE 2. Bacterial quality of fishes

Fish		Histamine content mg/100 g fish muscle				Histidine
FISA		1	2	3	4	decarboxylating bacteria/g
Jewfish (Otolithus) spp.		0,15	0.06	0,15	0,41	3,26 × 10*
Indian mackerel (Rastrelliger kanagurta)	••	0,20	0,14	0,42	0,29	2.18×10^{3}
Carangids (Chorinemus spp.)		0.88	0.25	0.81	1,30	4.40 × 10°
Malabar herring (Thrissocles sp.)	••	0,49	0,18	0	0	1.12 × 104
Silverbelly (Leiognathus spp.)		0,30	0,20	0	0	6.4 × 10°
Kilimeen (Nemipterus Japonicus)	••	0.18	0.16	0	0	196
Seerfish (Scomberomorus spp.)		0.53	0,38	0	0	40
Carangid	••	0	0	0	0	90
Lactarius lactarius		0.	0	0	0	3,85 × 10°
Ribbonfish (Trichiurus spp.)		0	0	0	0	2.0 × 10*

TABLE 3. Histamine content of individual fishes of each species in relation to their content of histidine decarboxylating bacteria

This observation is typical of fresh fish and fishes stored at low temperature (2 to 10°C) for periods 3 to 12 days (Salguero and Mackie, 1979; Pan and James, 1985; Ames et al., 1987). decarboxylating bacteria in all the samples histamine poisoning.

analysed. This shows that the bacterial activity had not been enough to degrade the histidine in fish muscle to histamine in sufficient quantities. Hence, it is assumed that fish sold in the However, there was incidence of histidine markets are safe from the point of view of

REFERENCES

ANON, 1987, Infofish Marketing Digest, 2: 38-39,

AMES, G. R., K. GOPAKUMAR AND P. K. VIJAYAN 1987. Ibid., 2:41.

DIFCO 1971. Microbiological and chemical laboratory procedures. 9th Edn. Difco Laboratorie's Inc. Detioit, Michigan.

FDA 1973, Bacteriological Analytical Mannual for Foods, Bureau of Foods, Food and Drug Administra-tion, U.S.A.

HARDY, R. and J. G. M. SMITH 1976. Journal of the Science of Food and Agriculture, 27: 595-599.

KIMATA, M. C. 1961. Fish as Food. Academ Press, New York, Chapter 10.

NEVEN, JR. C. F., M. B. JEFFREY AND D. A. CORLETT JR. 1981. Differential plating medium for quantitative detection of histamine producing bacteria. Appl Environ. Microbiol., 41: 321-322.

PAN, B. S. AND D. JAMES 1985. Histamine in marine products. Production by bacteria, measurement and prediction of formation. FAO Fisheries Technical Paper No. 252.

SALGUERO, J. F. AND I. M. MACKIE 1979. J. Fd. Technol., 14: 131-139.